

LADDL BACTERIOLOGY SPECIMEN SUBMISSION GUIDELINES

General Information:

1. Clearly label each sample container with name, date and sample site.
2. Best specimens are fluid and tissue collected and placed in sterile containers.
3. Swabs can be used, but typically only hold about 150 micro liters of fluid. Out of every 100 bacteria absorbed on a swab, only 3 make it to culture.
4. If a swab is used to collect the specimen, please use a culturette system that contains a buffered non-nutritive formula.
5. Enrichment broth, such as a blood culture bottle, can be used for transport and culture of joint fluid, CSF, or blood (typically sterile anatomical sites). Do not refrigerate. Leave at room temperature.
6. LADDL only accepts pond water specimens submitted by the Louisiana State Veterinarian's Office for *Vibrio cholerae* testing.

Aerobic Culture:

1. Swabs should be submitted in a transport media, such as a culturette system, to avoid drying of the swab and loss of bacteria. For best results, do not submit a dry, cotton swab.
2. Tissue should be taken as aseptically as possible and placed in a sterile container.
3. Avoid drying of tissue.
4. Biopsy Tissue can be transported in moist sterile gauze and placed in a sterile container.
5. Do not submit bacteriological specimens in an EDTA tube.
6. The best bacteriology results are obtained when specimens are transported to the lab as quickly as possible in the appropriate transport media with an adequate amount of material.
7. If direct exam and culture are requested, please send 2 swabs.
8. If identification of growth on blood agar plate is desired, please do not send plate. Instead, take sample of colony on culturette/swab and submit for identification.

Anaerobic Culture:

1. Specimens submitted for isolation of anaerobes require special transport media. Call the laboratory for further information.
2. Aerobic swabs from any site are generally unacceptable for anaerobic culture.
3. In general, specimens suitable for anaerobic culture are those that are collected by tissue biopsy or aspirated by needle and syringe.

Fecal Culture:

1. When submitting fecal or cloacal specimens, the following identification schemes are used:
 - 'Salmonella only' – media selected to recover only Salmonella
 - Routine fecal culture – media selected to recover enteric pathogens, such as *Salmonella*, *Yersinia*, *Aeromonas*, and *Plesiomonas*
 - Cloacal cultures are routinely set up for any Gram negative bacterium

- A specific request will be needed for *Campylobacter jejuni*, *Vibrio cholerae* or Johne's culturing.

Fungal Culture:

1. Tissues and swabs are the preferred samples for fungal culture.
2. All samples should be collected as aseptically as possible and stored in a sterile container.
3. Tissues, scales, hair, scabs, or scrapings collected from the center and edge of the lesion make ideal samples.
4. Fungal cultures are routinely conducted on chronic nasal discharges, aborted fetal tissues, tissue granulomas, and corneal/conjunctival specimens because of the prevalence of fungal infections at these sites.
5. Do not over inoculate vials of DTM with hair/scrapings. Plant only 2-3 diseased hairs.
6. Do not hold inoculated DTM cultures until the medium turns red. Bacteria and saprophytic fungi will each turn the medium red, so ship immediately.